

FORM PTO-1390 (Modified)
(REV 11-98)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

DEX-0105

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

09/700700

INTERNATIONAL APPLICATION NO.
PCT/US99/10548INTERNATIONAL FILING DATE
12 MAY 1999PRIORITY DATE CLAIMED
21 MAY 1998

TITLE OF INVENTION

A NOVEL METHOD OF DIAGNOSING, MONITORING, AND STAGING PROSTATE CANCER

APPLICANT(S) FOR DO/EO/US

ALL, Shujath et al.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ has been transmitted by the International Bureau.
 - c. ☒ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ A copy of the International Search Report (PCT/ISA/210).
8. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
10. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
11. ☐ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

Items 13 to 20 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ Certificate of Mailing by Express Mail
20. ☒ Other items or information:

Executed Verified Statement Claiming Small Entity Status

"Express Mail" Label No. #EL777534836US
Date of Deposit - November 20, 2000

I hereby certify that this paper is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 on the date indicated above and is addressed to the Assistant Commissioner of Patents, Box PCT, Washington, D.C. 20231.

By Suzanne Sparkman
Typed Name: Suzanne Sparkman

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.492 (a) (1) - (5)) : 09/700700	INTERNATIONAL APPLICATION NO. PCT/US99/10548	ATTORNEY'S DOCKET NUMBER DEX-0105
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21. The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :				CALCULATIONS PTO USE ONLY	
<input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1,000.00					
<input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00					
<input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00					
<input checked="" type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00					
<input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00					
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$690.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				\$0.00	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	11 - 20 =	0	x \$18.00	\$0.00	
Independent claims	5 - 3 =	2	x \$80.00	\$160.00	
Multiple Dependent Claims (check if applicable).			<input checked="" type="checkbox"/>	\$270.00	
TOTAL OF ABOVE CALCULATIONS =				\$1,120.00	
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) (check if applicable).			<input checked="" type="checkbox"/>	\$560.00	
SUBTOTAL =				\$560.00	
Processing fee of \$130.00 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492 (f)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				\$0.00	
TOTAL NATIONAL FEE =				\$560.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).			<input type="checkbox"/>	\$0.00	
TOTAL FEES ENCLOSED =				\$560.00	
				Amount to be: refunded	\$
				charged	\$

☐ A check in the amount of _____ to cover the above fees is enclosed.

☐ Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees.
A duplicate copy of this sheet is enclosed.

☒ **Credit Card Payment form attached.**

☒ The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. **501-619** A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

LICATA, Jane Massey Licata & Tyrrell P.C. 66 E. Main Street Marlton, New Jersey 08053 US	<u><i>Jane Massey Licata</i></u> SIGNATURE Jane Massey Licata NAME 32,257 REGISTRATION NUMBER 20 November 2000 DATE
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VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS (37 CFR 1.9(f) AND 1.27 (c)) - SMALL BUSINESS CONCERN			Docket No. DEX-0105
Serial No. Not Yet Assigned	Filing Date Herewith	Patent No.	Issue Date
Applicant/ ALI, Shujath et al. Patentee:			
Invention: A NOVEL METHOD OF DIAGNOSING, MONITORING, AND STAGING PROSTATE CANCER			
<p>I hereby declare that I am:</p> <p> <input type="checkbox"/> the owner of the small business concern identified below: <input checked="" type="checkbox"/> an official of the small business concern empowered to act on behalf of the concern identified below: </p> <p>NAME OF CONCERN: <u>diaDexus, Inc.</u></p> <p>ADDRESS OF CONCERN: <u>3303 Octavius Drive, Santa Clara, California 94043</u></p> <p>I hereby declare that the above-identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under Section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.</p> <p>I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the above identified invention described in:</p> <p> <input checked="" type="checkbox"/> the specification filed herewith with title as listed above. <input type="checkbox"/> the application identified above. <input type="checkbox"/> the patent identified above. </p> <p>If the rights held by the above-identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed on the next page and no rights to the invention are held by any person, other than the inventor, who could not qualify as an independent inventor under 37 CFR 1.9(c) or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).</p>			

Each person, concern or organization to which I have assigned, granted, conveyed, or licensed or am under an obligation under contract or law to assign, grant, convey, or license any rights in the invention is listed below:

- ☒ no such person, concern or organization exists.
☐ each such person, concern or organization is listed below.

FULL NAME _____

ADDRESS _____

☐

Individual

☐

Small Business Concern

☐

Nonprofit Organization

FULL NAME _____

ADDRESS _____

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Individual

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Small Business Concern

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Nonprofit Organization

FULL NAME _____

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Nonprofit Organization

FULL NAME _____

ADDRESS _____

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Individual

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Small Business Concern

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Nonprofit Organization

Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities. (37 CFR 1.27)

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING:

Mohan Iyer

TITLE OF PERSON SIGNING

OTHER THAN OWNER:

Vice President, Business Development

ADDRESS OF PERSON SIGNING:

diaDexus, Inc.

3303 Octavius Drive

Santa Clara, California 95054

SIGNATURE:

Mohan S. Iyer

DATE:

11/7/00

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A NOVEL METHOD OF DIAGNOSING,
MONITORING, AND STAGING PROSTATE CANCER

FIELD OF THE INVENTION

This invention relates, in part, to newly developed
5 assays for detecting, diagnosing, monitoring, staging, and
prognosticating cancers, particularly prostate cancer.

BACKGROUND OF THE INVENTION

Cancer of the prostate is the most prevalent malignancy in adult males, excluding skin cancer, and is an increasingly prevalent health problem in the United States. In 1996, it was estimated that in the United States, 41,400 deaths would result from this disease, indicating that prostate cancer is second only to lung cancer as the most common cause of death in the same population. If diagnosed and treated early, when the cancer is still confined to the prostate, the chance of cure is significantly higher.

Treatment decisions for an individual are linked to the stage of prostate cancer present in that individual. A common classification of the spread of prostate cancer was developed by the American Urological Association (AUA). The AUA classification divides prostate tumors into four stages, A to D. Stage A, microscopic cancer within prostate, is further subdivided into stages A1 and A2. Sub-stage A1 is a well-differentiated cancer confined to one site within the prostate. Treatment is generally observation, radical prostatectomy, or radiation. Sub-stage A2 is a moderately to poorly differentiated cancer at multiple sites within the prostate. Treatment is radical prostatectomy or radiation. Stage B, palpable lump within the prostate, is further subdivided into stages B1 and B2. In sub-stage B1, the cancer forms a small nodule in one lobe of the prostate. In sub-stage B2, the cancer forms large or multiple nodules, or occurs in both lobes of the prostate. Treatment for both sub-stages B1 and B2 is either radical prostatectomy or radiation.

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Stage C is a large cancer mass involving most or all of the prostate and is further subdivided into two stages. In sub-stage C1, the cancer forms a continuous mass that may have extended beyond the prostate. In sub-stage C2, the cancer
5 forms a continuous mass that invades the surrounding tissue. Treatment for both these sub-stages is radiation with or without drugs. The fourth stage is metastatic cancer and is also subdivided into two stages. In sub-stage D1, the cancer appears in the lymph nodes of the pelvis. In sub-stage D2,
10 the cancer involves tissues beyond lymph nodes. Treatment for both these sub-stages is systemic drugs to address the cancer as well as pain.

However, current prostate cancer staging methods are limited. As many as 50% of prostate cancers initially staged
15 as A2, B, or C are actually stage D, metastatic. Discovery of metastasis is significant because patients with metastatic cancers have a poorer prognosis and require significantly different therapy than those with localized cancers. The five year survival rates for patients with localized and metastatic
20 prostate cancers are 93% and 29%, respectively.

Accordingly, there is a great need for increasingly sensitive methods for the staging of a cancer in a human to determine whether or not such cancer has metastasized and for monitoring the progress of a cancer in a human.

25 In the present invention, methods are provided for detecting, diagnosing, monitoring, staging and prognosticating cancers, particularly prostate cancer via seven (7) Prostate Specific Genes (PSG). The seven PSGs refer, among other things, to native proteins expressed by the genes comprising
30 the polynucleotide sequences of any of SEQ ID NO: 1, 2, 3, 4, 5, 6 or 7. In the alternative, what is meant by the seven PSGs as used herein, means the native mRNAs encoded by the genes comprising any of the polynucleotide sequences of SEQ ID NO: 1, 2, 3, 4, 5, 6 or 7 or levels of the genes comprising

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any of the polynucleotide sequences of SEQ ID NO: 1, 2, 3, 4, 5, 6 or 7.

Other objects, features, advantages and aspects of the present invention will become apparent to those of skill in the art from the following description. It should be understood, however, that the following description and the specific examples, while indicating preferred embodiments of the invention, are given by way of illustration only. Various changes and modifications within the spirit and scope of the disclosed invention will become readily apparent to those skilled in the art from reading the following description and from reading the other parts of the present disclosure.

SUMMARY OF THE INVENTION

Toward these ends, and others, it is an object of the present invention to provide a method for diagnosing the presence of prostate cancer in a patient which comprises measuring levels of PSG in a sample of cells, tissue or bodily fluid from the patient and comparing the measured levels of PSG with levels of PSG in preferably the same cells, tissue, or bodily fluid type of a control, wherein an increase in the measured PSG levels in the patient versus levels of PSG in the control is associated with prostate cancer.

Another object of the present invention is to provide a method of diagnosing metastatic prostate cancer in a patient which comprises measuring PSG levels in a sample of cells, tissue, or bodily fluid from the patient and comparing the measured PSG levels with levels of PSG in preferably the same cells, tissue, or bodily fluid type of a control, wherein an increase in measured PSG levels in the patient versus levels of PSG in the control is associated with a cancer which has metastasized.

Another object of the present invention is to provide a method of staging prostate cancer in a patient which comprises identifying a patient having prostate cancer,

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understood, however, that the following description and the specific examples, while indicating preferred embodiments of the invention, are given by way of illustration only. Various changes and modifications within the spirit and scope of the disclosed invention will become readily apparent to those skilled in the art from reading the following description and from reading the other parts of the present disclosure.

DESCRIPTION OF THE INVENTION

The present invention relates to diagnostic assays and methods, both quantitative and qualitative for detecting, diagnosing, monitoring, staging, and prognosticating cancers by comparing levels of PSG measured in a patient with levels of PSG in a control. What is meant by "levels of PSG" as used herein, means levels of the native protein expressed by the gene comprising the polynucleotide sequence of any of SEQ ID NO: 1, 2, 3, 4, 5, 6 or 7. In the alternative, what is meant by "levels of PSG" as used herein, is levels of the native mRNA encoded by the gene comprising any of the polynucleotide sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6 or 7 or levels of the gene comprising any of the polynucleotide sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6 or 7. Such levels are preferably measured in at least one of cells, tissues and/or bodily fluids, and includes determination of both normal and abnormal levels of PSGs. Thus, for instance, a diagnostic assay in accordance with the invention for diagnosing overexpression of PSG protein compared to control bodily fluids, cells, or tissue samples may be used to diagnose the presence of cancers, including prostate cancer. Any of the seven PSGs may be measured alone in the methods of the invention, all together or in various combinations of the seven PSGs.

By "control" it is meant a human patient without cancer and/or non cancerous samples from the patient, also referred to herein as a normal human control; in the methods for diagnosing or monitoring for metastasis, control may also

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include samples from a human patient that is determined by reliable methods to have prostate cancer which has not metastasized.

All the methods of the present invention may optionally include measuring the levels of other cancer markers as well as PSG. Other cancer markers, in addition to PSG, useful in the present invention will depend on the cancer being tested and are known to those of skill in the art. For example, simultaneous testing for increases in PSA as well as increases in PSG are also within the scope of the present invention and believed to provide a higher level of assurance that such cancer being tested is metastatic or the onset of metastasis has occurred.

Diagnostic Assays

The present invention provides methods for diagnosing the presence of prostate cancer by analyzing for changes in levels of PSG in cells, tissues or bodily fluids compared with levels of PSG in cells, tissues or bodily fluids of preferably the same type from a normal human control, wherein an increase in levels of PSG in the patient versus the normal human control is associated with the presence of prostate cancer. Without limiting the instant invention, typically, for a quantitative diagnostic assay a positive result indicating the patient being tested has cancer is one in which cells, tissues, or bodily fluid levels of the cancer marker, such as PSG, are at least two times higher, and most preferably are at least five times higher, than in preferably the same cells, tissues, or bodily fluid of a normal human control.

The present invention also provides a method of diagnosing metastatic prostate cancer in a patient having prostate cancer which has not yet metastasized for the onset of metastasis. In the method of the present invention, a human cancer patient suspected of having prostate cancer which may have metastasized (but which was not previously known to have metastasized) is identified. This is accomplished by a

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variety of means known to those of skill in the art. For example, in the case of prostate cancer, patients are typically diagnosed with prostate cancer following traditional detection methods.

5 In the present invention, determining the presence of PSG in cells, tissues, or bodily fluid, is particularly useful for discriminating between prostate cancer which has not metastasized and prostate cancer which has metastasized.

Existing techniques have difficulty discriminating
10 between prostate cancer which has metastasized and prostate cancer which has not metastasized and proper treatment selection is often dependent upon such knowledge.

In the present invention, the cancer marker levels measured in such cells, tissue, or bodily fluid are PSGs, and
15 are compared with levels of PSG in preferably the same cells, tissue, or bodily fluid type of a normal human control. That is, if the cancer marker being observed is just PSG in serum, this level is preferably compared with the level of PSG in serum of a normal human patient. An increase in the PSG in
20 the patient versus the normal human control is associated with prostate cancer which has metastasized.

Without limiting the instant invention, typically, for a quantitative diagnostic assay a positive result indicating the cancer in the patient being tested or monitored
25 has metastasized is one in which cells, tissues, or bodily fluid levels of the cancer marker, such as PSG, are at least two times higher, and most preferable are at least five times higher, than in preferably the same cells, tissues, or bodily fluid of a normal patient.

30 **Staging**

The invention also provides a method of staging prostate cancer in a human patient.

The method comprises identifying a human patient having such cancer and analyzing a sample of cells, tissues,
35 or bodily fluid from such patient for PSG. Then, the method

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compares PSG levels in such cells, tissues, or bodily fluid with levels of PSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein an increase in PSG levels in the patient versus the normal human control is associated with a cancer which is progressing and a decrease in the levels of PSG is associated with a cancer which is regressing or in remission.

Monitoring

Further provided is a method of monitoring prostate cancer in a human having such cancer for the onset of metastasis. The method comprises identifying a human patient having such cancer that is not known to have metastasized; periodically analyzing a sample of cells, tissues, or bodily fluid from such patient for PSG; and comparing the PSG levels in such cells, tissue, or bodily fluid with levels of PSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein an increase in PSG levels in the patient versus the normal human control is associated with a cancer which has metastasized.

Further provided by this invention is a method of monitoring the change in stage of prostate cancer in a human having such cancer. The method comprises identifying a human patient having such cancer; periodically analyzing a sample of cells, tissue, or bodily fluid from such patient for PSG; comparing the PSG levels in such cells, tissue, or bodily fluid with levels of PSG in preferably the same patient.

Monitoring such patient for onset of metastasis is periodic and preferably done on a quarterly basis. However, this may be more or less frequent depending on the cancer, the particular patient, and the stage of the cancer.

Assay Techniques

Assay techniques that can be used to determine levels of gene expression, such as PSG of the present invention, in a sample derived from a host are well-known to those of skill in the art. Such assay methods include radioimmunoassays,

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reverse transcriptase PCR (RT-PCR) assays, immunohistochemistry assays, *in situ* hybridization assays, competitive-binding assays, Western Blot analyses and ELISA assays. Among these, ELISAs are frequently preferred to
5 diagnose a gene's expressed protein in biological fluids. An ELISA assay initially comprises preparing an antibody, if not readily available from a commercial source, specific to PSG, preferably a monoclonal antibody. In addition a reporter antibody generally is prepared which binds specifically to
10 PSG. The reporter antibody is attached to a detectable reagent such as radioactive, fluorescent or enzymatic reagent, for example horseradish peroxidase enzyme or alkaline phosphatase.

To carry out the ELISA, antibody specific to PSG is
15 incubated on a solid support, e.g., a polystyrene dish, that binds the antibody. Any free protein binding sites on the dish are then covered by incubating with a non-specific protein such as bovine serum albumin. Next, the sample to be analyzed is incubated in the dish, during which time PSG binds
20 to the specific antibody attached to the polystyrene dish. Unbound sample is washed out with buffer. A reporter antibody specifically directed to PSG and linked to horseradish peroxidase is placed in the dish resulting in binding of the reporter antibody to any monoclonal antibody bound to PSG.
25 Unattached reporter antibody is then washed out. Reagents for peroxidase activity, including a colorimetric substrate are then added to the dish. Immobilized peroxidase, linked to PSG antibodies, produces a colored reaction product. The amount of color developed in a given time period is proportional to
30 the amount of PSG protein present in the sample. Quantitative results typically are obtained by reference to a standard curve.

A competition assay may be employed wherein antibodies specific to PSG attached to a solid support and
35 labeled PSG and a sample derived from the host are passed over

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the solid support and the amount of label detected attached to the solid support can be correlated to a quantity of PSG in the sample.

Nucleic acid methods may be used to detect PSG mRNA as a marker for prostate cancer. Polymerase chain reaction (PCR) and other nucleic acid methods, such as ligase chain reaction (LCR) and nucleic acid sequence based amplification (NASABA), can be used to detect malignant cells for diagnosis and monitoring of various malignancies. For example, reverse-transcriptase PCR (RT-PCR) is a powerful technique which can be used to detect the presence of a specific mRNA population in a complex mixture of thousands of other mRNA species. In RT-PCR, an mRNA species is first reverse transcribed to complementary DNA (cDNA) with use of the enzyme reverse transcriptase; the cDNA is then amplified as in a standard PCR reaction. RT-PCR can thus reveal by amplification the presence of a single species of mRNA. Accordingly, if the mRNA is highly specific for the cell that produces it, RT-PCR can be used to identify the presence of a specific type of cell.

Hybridization to clones or oligonucleotides arrayed on a solid support (i.e., gridding) can be used to both detect the expression of and quantitate the level of expression of that gene. In this approach, a cDNA encoding the PSG gene is fixed to a substrate. The substrate may be of any suitable type including but not limited to glass, nitrocellulose, nylon or plastic. At least a portion of the DNA encoding the PSG gene is attached to the substrate and then incubated with the analyte, which may be RNA or a complementary DNA (cDNA) copy of the RNA, isolated from the tissue of interest.

Hybridization between the substrate bound DNA and the analyte can be detected and quantitated by several means including but not limited to radioactive labeling or fluorescence labeling of the analyte or a secondary molecule designed to detect the hybrid. Quantitation of the level of

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gene expression can be done by comparison of the intensity of the signal from the analyte compared with that determined from known standards. The standards can be obtained by *in vitro* transcription of the target gene, quantitating the yield, and
5 then using that material to generate a standard curve.

The above tests can be carried out on samples derived from a variety of patients' cells, bodily fluids and/or tissue extracts (homogenates or solubilized tissue) such as from tissue biopsy and autopsy material. Bodily fluids useful in
10 the present invention include blood, urine, saliva, or any other bodily secretion or derivative thereof. Blood can include whole blood, plasma, serum, or any derivative of blood.

EXAMPLES

15 The present invention is further described by the following examples. These examples are provided solely to illustrate the invention by reference to specific embodiments. These exemplifications, while illustrating certain specific aspects of the invention, do not portray the limitations or
20 circumscribe the scope of the disclosed invention.

EXAMPLE 1: PSGs

Searches were carried out and PSGs identified using the following Search Tools as part of the LIFESEQ® database available from Incyte Pharmaceuticals, Palo Alto, CA:

25 1. Library Comparison (compares one library to one other library) allows the identification of clones expressed in tumor and absent or expressed at a lower level in normal tissue.

30 2. Subsetting is similar to library comparison but allows the identification of clones expressed in a pool of libraries and absent or expressed at a lower level in a second pool of libraries.

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3. Transcript Imaging lists all of the clones in a single library or a pool of libraries based on abundance. Individual clones can then be examined using Electronic Northern

5 ESTs.

4. Protein Function: Incyte has identified subsets of ESTs with a potential protein function based on homologies to known proteins. Some examples in this database include Transcription Factors and Proteases. Some leads were

10 identified by searching in this database for clones whose component ESTs showed disease specificity.

Electronic subtractions, transcript imaging and protein function searches were used to identify clones, whose component ESTs were exclusively or more frequently found in

15 libraries from specific tumors. Individual candidate clones were examined in detail by checking where each EST originated.

Table 1:

SEQ ID NO:	Clone ID #	Gene ID #	
1	1550426	244673	Protein Function (Transcription Factors)
2	1255804	14878	Subsetting
3	1808432	255819	Subsetting
4	3930803	none	Subsetting
5	645804	235032	Subsetting
6	1862352	221558	Subsetting
7	1450626	236019	Subsetting

**EXAMPLE 2: Measurement of SEQ ID NO:1; Clone ID # 1550426;
Gene ID #244673 (pro101)**

30 The example is carried out using standard techniques, which are well known and routine to those of skill in the art,

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except where otherwise described in detail. Routine molecular biology techniques of the following example are carried out as described in standard laboratory manuals, such as Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989).

Relative Quantitation of Gene Expression

Real-time quantitative PCR with fluorescent Taqman probes is a quantitative detection system utilizing the 5'-3' nuclease activity of Taq DNA polymerase. The method uses an internal fluorescent oligonucleotide probe (Taqman) labeled with a 5' reporter dye and a downstream, 3' quencher dye. During PCR, the 5'-3' nuclease activity of Taq DNA polymerase releases the reporter, whose fluorescence can then be detected by the laser detector of the Model 7700 Sequence Detection System (PE Applied Biosystems, Foster City, CA, USA).

Amplification of an endogenous control is used to standardize the amount of sample RNA added to the reaction and normalize for Reverse Transcriptase (RT) efficiency. Either cyclophilin, glyceraldehyde-3-phosphate dehydrogenase (GAPDH) or 18S ribosomal RNA (rRNA) is used as this endogenous control. To calculate relative quantitation between all the samples studied, the target RNA levels for one sample are used as the basis for comparative results (calibrator). Quantitation relative to the "calibrator" is obtained using the standard curve method or the comparative method (User Bulletin #2: ABI PRISM 7700 Sequence Detection System).

To evaluate the tissue distribution, and the level of prol01 (SEQ ID NO:1) in normal and tumor tissue, total RNA was extracted from tumor and matched normal adjacent tissues and from unmatched tumor and normal tissues. Subsequently, first strand cDNA was prepared with reverse transcriptase and the polymerase chain reaction carried out using primers and Taqman probe specific to prol01 (SEQ ID NO:1). The results

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were obtained using the ABI PRISM 7700 Sequence Detector. The absolute numbers are relative levels of expression of pro101 (SEQ ID NO:1) compared to the calibrator.

The absolute numbers are depicted in the following Table 2 as relative levels of expression in 12 normal tissues of pro101 (SEQ ID NO:1) compared to kidney (calibrator). These RNA samples were generated by pooling samples from a particular tissue from different individuals.

Table 2: Relative levels of pro101 Expression in Pooled Samples

Tissue	NORMAL
Brain	1.2
Heart	2
Kidney	1
Liver	7.2
Lung	48.2
Mammary	2.5
Prostate	1418.4
Spleen	1.6
Small	1.9
Testis	57.3
Thymus	1.3
Uterus	7.6

The relative levels of expression in Table 2 show that for the PSG pro101 (SEQ ID NO:1) mRNA expression is more than 20 fold higher in the pool of normal prostate compared with the other 11 normal tissue pools analyzed. These results demonstrate that mRNA expression of the PSG is highly specific for prostate.

The tissues shown in Table 2 correspond to pools of samples from different individuals. The tissues shown in the following Table 3 were obtained from individuals and are not pooled. Hence the values for mRNA expression levels shown in Table 2 cannot be directly compared to the values shown in Table 3.

- 15 -

The absolute numbers in Table 3 are relative levels of expression of pro101 (SEQ ID NO:1) compared to kidney (calibrator), in 60 pairs of matching samples. Each matching pair contains the cancer sample for a particular tissue and the normal adjacent sample for that same tissue from the same individual. The results from 3 unmatched ovary tumor, 3 unmatched normal ovary, 1 unmatched mammary tumor and 1 unmatched normal mammary gland are also shown.

Table 3: Relative Levels of pro101 Expression in Individual Samples

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TISSUE	CANCER	MATCHING	UNMATCHED
Prostate 1	103.9	0	
Prostate 2	2219	84.2	
Prostate 3	5048.2	3623.6	
Prostate 4	11052.3	2029.4	
Prostate 5	229.1	41.1	
Prostate 6	57.9	25.3	
Prostate 7	58.5	57.069	
Prostate 8	1074.6	610.8	
Prostate 9	32.7	79.3	
Prostate 10	15.8	2.09	
Prostate 11	436.4	438	
Prostate 12	49.5	59.3	
Prostate 13	128	56	
Bladder 1	0	0	
Bladder 2	0	0	
Bladder 3	0.7	0	
Colon 1	0	0	
Colon 2	0	0	
Colon 3	0	0	
Colon 4	3.3	1.9	
Colon 5	0.1	0.8	
Colon 6	0	0	
Lung 1	0	0	
Lung 2	0.5	1.6	
Lung 3	1.4	2.1	
Lung 4	0	0	
Lung 5	0	0	
Kidney 1	0	0	
Kidney 2	0	0	
Kidney 3	0	0	
Kidney 4	0	0	
Liver 1	1.5	5.7	
Liver 2	26.9	7.9	
Liver 3	0	0	

- 16 -

	Pancreas 1	0.9	0.9	
	Pancreas 2	3	0	
	Pancreas 3	0	0	
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	Mammary 6			0
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	Ovary 3	1.5		
	Ovary 4			22.9
	Ovary 5	0		
	Ovary 6			1.8

35 Among 128 samples in Table 3 representing 14 different tissues, the higher levels of expression are consistently in prostate tissues. These results confirm the tissue specificity results obtained with normal samples shown in Table 2. Table 2 and Table 3 represent a combined total of

40 140 samples in 18 human tissue types. Sixty-eight samples representing 13 different tissue types excluding prostate had no detected prol01 mRNA (Table 3). In 4 tissues (stomach small intestine kidney and testis) no prol01 (SEQ ID NO:1) mRNA was detected for any sample tested from individuals

45 (Table 3). Expression of this PSG was detected in testis in the pooled normal sample (Table 3). The median expression in

- 17 -

prostate cancer samples in Table 3 is 166.5 units. Excluding Ovary 4 (Normal), only 1 sample in Table 3, Liver 2 (Cancer), is greater than 10% of this value.

Comparisons of the level of mRNA expression in prostate tumor samples and the normal adjacent tissue from the same individuals are also shown in Table 3. The PSG pro101 (SEQ ID NO:1) is expressed at higher levels in 9 of 13 (69%) prostate cancer tissues (Prostate 1, 2, 3, 4, 5, 6, 8, 10 and 13) compared with the corresponding normal adjacent tissue. The level of expression of this PSG is lower in prostate tumor compared to normal adjacent tissue in two samples (Prostate 9 and 12). Equivalent levels of expression were detected in two matched samples (Prostate 7 and 11). Previous mRNA expression analysis for genes coding for the diagnostic markers PSA and PLA2 showed higher expression of the mRNA in 40% to 80% of the tumor samples compared to matching normal adjacent tissue. Higher expression in the tumor sample compared to the corresponding normal adjacent tissue is observed for Bladder 3, Colon 4, Liver 2, Pancreas 2, Endometrium 5 and. Mammary 1, 2 and 3. Higher expression in the normal adjacent samples is observed for Colon 5, Lung 2, Lung 3, Liver 1, Endometrium 1 and Uterus 4. However, the levels detected are in most cases comparable amongst the different tissues and low compared to levels found in most prostate tissues.

The high level of tissue specificity, plus the mRNA overexpression in 9 of 13 of the prostate tumor samples tested compared to the normal adjacent tissues are believed to make the PSG, pro101 (SEQ ID NO:1) a good diagnostic marker for detection of prostate cancer using mRNA.

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What is Claimed is:

1. A method for diagnosing the presence of prostate cancer in a patient comprising:

(a) measuring levels of PSG in a sample of cells, tissue or bodily fluid obtained from the patient; and

(b) comparing the measured levels of PSG with levels of PSG in a sample of cells, tissue or bodily fluid obtained from a control, wherein an increase in measured levels of PSG in the patient versus the PSG levels in the control is associated with the presence of prostate cancer.

2. A method of diagnosing metastatic prostate cancer in a patient comprising:

(a) measuring levels of PSG in a sample of cells, tissue, or bodily fluid obtained from the patient; and

(b) comparing the measured levels of PSG with levels of PSG in a sample of cells, tissue, or bodily fluid obtained from a control, wherein an increase in measured PSG levels in the patient versus the PSG levels in the control is associated with a cancer which has metastasized.

3. A method of staging prostate cancer in a patient comprising:

(a) identifying a patient suffering from prostate cancer;

(b) measuring levels of PSG in a sample of cells, tissue, or bodily fluid obtained from the patient; and

(c) comparing the measured levels of PSG with levels of PSG in a sample of cells, tissue, or bodily fluid obtained from a control, wherein an increase in the measured levels of PSG versus the levels of PSG in the control is associated with a cancer which is progressing and a decrease in the measured levels of PSG versus the levels of PSG in the control is associated with a cancer which is regressing or in remission.

(a) identifying a patient having prostate cancer that is not known to have metastasized;

(c) comparing the periodically measured levels of PSG with levels of PSG in cells, tissue, or bodily fluid obtained from a control, wherein an increase in any one of the periodically measured levels of PSG in the patient versus the levels of PSG in the control is associated with a cancer which has metastasized.

15 (a) identifying a patient having prostate cancer;
 (b) periodically measuring levels of PSG in samples
of cells, tissue, or bodily fluid obtained from the patient;
and

6. The method of claim 1, 2, 3, 4 or 5 wherein the PSG comprises SEQ ID NO:1.

Docket No.

DEX-0105

Declaration and Power of Attorney For Patent Application

English Language Declaration

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

A NOVEL METHOD OF DIAGNOSING, MONITORING, AND STAGING PROSTATE CANCER

the specification of which

(check one)

☐ is attached hereto.

☒ was filed on 12 May 1999 as United States Application No. or PCT International

Application Number PCT/US99/10548

and was amended on _____

(if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, Section 119(a)-(d) or Section 365(b) of any foreign application(s) for patent or inventor's certificate, or Section 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Priority Not Claimed

(Number)

(Country)

(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional application(s) listed below:

60/086,265

21 May 1998

(Application Serial No.)

(Filing Date)

(Application Serial No.)

(Filing Date)

(Application Serial No.)

(Filing Date)

I hereby claim the benefit under 35 U. S. C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, C. F. R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)
(patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. *(list name and registration number)*



26259

PATENT TRADEMARK OFFICE

Send Correspondence to:

Direct Telephone Calls to: *(name and telephone number)*

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Sole or first inventor's signature	Date
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Fifth inventor's signature	Date
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Full name of sixth inventor, if any	
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